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G2P PCR Cleanup and Gel Extraction combo kit

Cat # PUR14-50
Pack Size: Suitable for 50 rxn.

Introduction:

The G2P PCR cleanup and Gel Extraction combo kit offers PCR Cleanup and gel elution in the same kit, very handy for cloning projects. Utilizing proprietary silica-based membrane technology, the kit delivers high yields of pure DNA fragments in a simple and rapid protocol that takes less time to complete. The kit is compatible with a wide range of downstream applications and is backed by a team of experienced technical support specialists.

Kit Content

- Buffer PCA (For PCR Cleanup)- 30mL
- Buffer GE (Gel dissolution and DNA Binding Buffer: For gel extraction) - 30mL
- Buffer D (Wash Buffer) * **Add 40ml of absolute ethanol before use.** (Common buffer) - 10mL
- Buffer E (Elution Buffer) (Common buffer) -5mL
- Binding Columns (50 Nos) and Collection Tubes (50 Nos)

Procedure for PCR Cleanup

1. Mix PCA in 1:1 (Vol:Vol) with PCR product
2. Load upto 600ul on Spin Column and spin at high speed (10,000xg) and load the supernatant on the spin column.
3. *Discard the flow-through.*
4. *Add 600 µl of Buffer D and spin for 3 minutes.*
5. *Discard the flow-through.*
6. *Spin again for 3 minutes to dry.*
7. *Place the column on a fresh microfuge tube (not included in the kit) and add approximately 30 µl of Buffer E. Water may be used in place of Buffer E for elution. When using water, ensure that the pH of the water is 8.0. When using lower volumes of elution buffer, add the buffer dropwise to the center of the membrane. Wait for 1 minute after adding the Buffer E.*
8. *Spin at high speed and collect the elution (gel-extracted product).*

Note: It is highly recommended to use low melting agarose or use a low percentage gel (~0.8%) for better recovery.

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Procedure For Gel Extraction

1. *Excise the gel band containing the DNA fragment using a clean, sharp scalpel. Remove any excess agarose from the sides of the gel band.*
2. *Weigh the gel slice in a microfuge tube and add 3 volumes of Buffer GE to 1 volume of gel (approximately 300 µl of Buffer GE to 100 mg of gel).*
3. *Incubate the mixture at 50°C for 10 minutes or until the gel slice is completely dissolved. You may shake the tube or vortex it lightly, or pipette mix. If using higher percentage gels (>2%), increase the incubation time.*
4. *Load up to 600 µl of the mixture onto a spin column and spin at high speed (10,000xg). Load the leftover solution and repeat this step if necessary.*
5. *Discard the flow-through.*
6. *Add 600 µl of Buffer D and spin for 3 minutes.*
7. *Discard the flow-through.*
8. *Spin again for 3 minutes to dry.*
9. *Place the column on a fresh microfuge tube (not included in the kit) and add approximately 30 µl of Buffer E. Water may be used in place of Buffer E for elution. When using water, ensure that the pH of the water is 8.0. When using lower volumes of elution buffer, add the buffer dropwise to the center of the membrane. Wait for 1 minute after adding the Buffer E.*
10. *Spin at high speed and collect the elution (gel-extracted product).*

Note: It is highly recommended to use low melting agarose or use a low percentage gel (~0.8%) for better recovery.